



Colorimetric Hydrogen Peroxide Assay Kit

Product Information

Cat

Kit-0990

Common Name

H₂O₂

Cat.No.

Kit-0990

Product Overview

This Colorimetric Hydrogen Peroxide Assay Kit uses our unique IR peroxidase substrate to quantify hydrogen peroxide in solutions and cell extracts. Upon hydrogen peroxide oxidation the colorless IR generates an intense blue color product that is pH-independent from pH 4 to 10. The existing colorimetric hydrogen peroxide assays (from other vendors) often have severe sample interferences caused by the inherent absorption of biological samples. The near infrared absorption of IR product minimizes the assay background that since the biological samples rarely absorb light beyond 600 nm. It can also be used to detect a variety of oxidase activities through enzyme-coupled reactions. The kit is an optimized 'mix and read' assay that is compatible with HTS liquid handling instruments.

Description

Hydrogen peroxide (H₂O₂) is a reactive oxygen metabolic by-product that serves as a key regulator for numerous oxidative stress-related states. It is involved in a number of biological events that have been linked to asthma, atherosclerosis, diabetic vasculopathy, osteoporosis, a number of neurodegenerative diseases and Down's syndrome. Perhaps the most intriguing aspect of H₂O₂ biology is the recent report that antibodies have the capacity to convert molecular oxygen into H₂O₂ to contribute to the normal recognition and destruction processes of the immune system. Measurement of this reactive species will help to determine how oxidative stress modulates varied intracellular pathways.

Storage

Keep in freezer and avoid exposure to light.



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Size

500 Tests

Kit Components

Component A: IR Peroxidase Substrate 1 vial

Component B: H₂O₂ 1 vial (3% stabilized solution, 200 μ L)

Component C: Assay Buffer 1 bottle (100 mL)

Component D: Horseradish Peroxidase 1 vial (20 units)

Component E: DMSO 1 vial (0.5 mL)

Preparation

1. Prepare stock solutions:

1.1 100X IR peroxidase substrate stock solution: Add 250 μ L of DMSO (Component E) into the vial of IR Peroxidase Substrate (Component A). The stock solution should be used promptly; any remaining solution should be aliquoted and refrozen at -20°C.

Note: Avoid repeated freeze-thaw cycles, and protect from light.

1.2 20 U/mL peroxidase stock solution: Add 1 mL of Assay Buffer (Component C) into the vial of Horseradish Peroxidase (Component D).

Note: The unused HRP solution should be divided into single use aliquots and stored at -20°C.

1.3 20 mM H₂O₂ stock solution: Add 22.7 μ L of 3% H₂O₂ (0.88 M, Component B) into 977 μ L of Assay Buffer (Component C).

Note: The diluted H₂O₂ stock solution is not stable. The unused portion should be discarded.

2. Prepare H₂O₂ reaction mixture: Prepare the H₂O₂ reaction mixture according to the following table and keep from light.

Table 1. H₂O₂ Reaction mixture for one 96-well plate (2X)

100X IR Peroxidase Substrate Stock Solution (from Step 1.1) 50 μ L

20 U/mL Peroxidase Stock Solution (from Step 1.2) 200 μ L

Assay Buffer (Component C) 4.75 mL

Total volume 5 mL

3. Prepare serial dilutions of H₂O₂ standard (0 to 50 μ M):

Warning 1: IR Peroxidase Substrate (Component A) is unstable in the presence of thiols such as DTT



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and β -mercaptoethanol. If the final concentration of the thiols is higher than 10 μ M, it would significantly decrease the assay dynamic range.

Warning 2: NADH and glutathione (reduced form of GSH) may interfere with the assay.

3.1 Add 5 μ L of 20 mM H₂O₂ stock solution (from Step 1.3) into 995 μ L of Assay Buffer (Component C) to get 100 μ M H₂O₂ standard.

3.2 Take 200 μ L of 100 μ M H₂O₂ standard to perform 1:2 serial dilutions to get 50, 25, 12.5, 6.25, 3.125, 1.56 and 0 μ M serial dilutions of H₂O₂ standard.

3.3 Add serial dilutions of H₂O₂ standard and H₂O₂-containing test samples into a white wall/clear bottom 96-well microplate as described in Tables 2 and 3.

Table 2 Layout of H₂O₂ standards and test samples in a white wall/clear bottom 96-well microplate

BL BL TS TS

HS1 HS1

HS2 HS2

HS3 HS3

HS4 HS4

HS5 HS5

HS6 HS6

HS7 HS7

Note: HS= H₂O₂ Standards; BL=Blank Control; TS=Test Samples

Table 3 Reagent composition for each well

H₂O₂ Standard: Serial Dilutions*: 50 μ L

Blank Control: Assay Buffer (Component C): 50 μ L

Test Sample: 50 μ L

*Note: Add the serial diluted of H₂O₂ standard from 1.56 μ M to 50 μ M into wells from HS1 to HS7 in duplicate.

Assay Protocol

4. Run H₂O₂ assay in supernatants reaction:

4.1 Add 50 μ L of H₂O₂ reaction mixture (from Step 2) into each well of H₂O₂ standard, blank control, and test samples (see Step 3.3) to make the total volume of 100 μ L/well.

Note: For a 384-well plate, add 25 μ L of sample and 25 μ L of H₂O₂ reaction mixture into



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each well.

4.2 Incubate the reaction at room temperature for 10 to 30 minutes, protected from light.

4.3 Monitor the absorbance with an absorbance plate reader at 650 nm.

5. Run H₂O₂ assay for cells:

The Colorimetric Hydrogen Peroxide Assay Kit can be used to measure the release of H₂O₂ from cells. The following is a suggested protocol that can be modified to meet the specific research needs.

5.1 The H₂O₂ reaction mixture should be prepared as Step 2 except that the Assay Buffer (Component C) should be replaced with the media used in your cell culture system. Suggested media including (a) Krebs Ringers Phosphate Buffer (KRPB); (b). Hanks Balanced Salt Solution (HBSS); or (c) Serum-free media.

5.2 Prepare cells in a 96-well plate (50 - 100 μ L/well), and activate the cells as desired.

Note: The negative controls (media alone and non-activated cells) are included for measuring the background fluorescence.

5.3 Add 50 μ L of H₂O₂ reaction mixture (from Step 5.1) into each well of cells, and H₂O₂ standards (from Step 3.3).

Note: For a 384-well plate, add 25 μ L of cells and 25 μ L of H₂O₂ reaction mixture into each well.

5.4 Incubate the reaction at room temperature for 10 to 60 minutes, protected from light.

5.5 Monitor the absorbance with an absorbance plate reader at 650 nm.

Analysis

The absorbance in blank wells (with the assay buffer only) is used as a control, and is subtracted from the values for those wells with the HRP reactions.
