

BCA Protein Quantification Kit (Enhanced)

Cat. No. Kit-0867 **Lot. No.** (See product label)

SPECIFICATION

Product Overview

Description BCA Protein Assay Kit is a ready-to-use detergent-compatible Western blot related total protein analysis reagent used for the quick determination of total protein concentration by measuring absorbance at 562 nm and comparing to a protein standard absorption vs. concentration curve, according to Smith.

Applications Western blotting, protein expression assays, protein profiling and characterization, protein quantitation assays

Usage Studying protein-protein interactions, Measuring column fractions after affinity chromatography, Assessing protein yields in whole cell lysates, High-throughput screening of fusion proteins

Storage Store at room temperature for one year.

Size Tube procedure: 50 assays
Microplate procedure: 500 assays

Kit Components BCA Reagent A: 100 ml
BCA Reagent B: 5 ml
Albumin (BSA) Standards: 20 ml (2 mg/ml)

Materials Required but Not Supplied Test tubes or microplates; Spectrophotometer; Microplate Reader

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Detection method	Colorimetric/Spectroscopic																																								
Compatible Sample Types	Serum, Plasma, Cell culture extracts, Tissue Extracts																																								
Assay Protocol	A. Test Tube Procedure 1. Mix BCA Reagent A and BCA Reagent B in the ratio of 50:1. i.e., mix 50 ml of BCA Reagent A with 1 ml BCA Reagent B. 2. Follow Table 1 to prepare a fresh set of standards. (Dilute Albumin (BSA) Standards with 0.9% NaCl or PBS) Table 1: Preparation of Albumin (BSA) Standards 3. Add 0.1 ml of each standard and protein samples into separate labeled test tubes. <table><thead><tr><th>Tube Number</th><th>Volume of Diluent (μl)</th><th>Volume of BSA (μl)</th><th>Final BSA Concentration (μg/ml)</th></tr></thead><tbody><tr><td>A</td><td>0 μl</td><td>900 μl of 2 mg/ml Stock</td><td>2000 μg/ml</td></tr><tr><td>B</td><td>100 μl</td><td>300 μl of tube A</td><td>1500 μg/ml</td></tr><tr><td>C</td><td>300 μl</td><td>300 μl of tube A</td><td>1000 μg/ml</td></tr><tr><td>D</td><td>200 μl</td><td>200 μl of tube B</td><td>750 μg/ml</td></tr><tr><td>E</td><td>300 μl</td><td>300 μl of tube C</td><td>500 μg/ml</td></tr><tr><td>F</td><td>300 μl</td><td>300 μl of tube E</td><td>250 μg/ml</td></tr><tr><td>G</td><td>300 μl</td><td>300 μl of tube F</td><td>125 μg/ml</td></tr><tr><td>H</td><td>400 μl</td><td>100 μl of tube G</td><td>25 μg/ml</td></tr><tr><td>I</td><td>300 μl</td><td>0 0 (blank)</td><td></td></tr></tbody></table> 4. Add 2 ml of BCA working reagent to each tube and mix well. 5. Incubate at 37°C for 30 minutes. Note: Increasing the incubation time and temperature can increase the net 562 nm absorbance for each test and decreases both minimum detection level and test range of the kit. 6. Cool all tubes to room temperature (RT). 7. Set the wavelength of spectrophotometer at OD 562 nm. Calibrate the instrument	Tube Number	Volume of Diluent (μl)	Volume of BSA (μl)	Final BSA Concentration (μg/ml)	A	0 μl	900 μl of 2 mg/ml Stock	2000 μg/ml	B	100 μl	300 μl of tube A	1500 μg/ml	C	300 μl	300 μl of tube A	1000 μg/ml	D	200 μl	200 μl of tube B	750 μg/ml	E	300 μl	300 μl of tube C	500 μg/ml	F	300 μl	300 μl of tube E	250 μg/ml	G	300 μl	300 μl of tube F	125 μg/ml	H	400 μl	100 μl of tube G	25 μg/ml	I	300 μl	0 0 (blank)	
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I	300 μl	0 0 (blank)																																							

to zero by using water. Subsequently, measure the absorbance of all samples within 10 minutes.

Note: Color development continues even after cooling to RT. However, the subsequent development at RT is too weak to produce significant error if all absorbance measurements are made within 10 minute.

8. Subtract OD562 of Blank from all readings.

9. Plot the BSA standard curve: OD562 (on Y axis) vs BSA Standard concentration (on X axis). Use the standard curve to determine the protein concentration of each unknown sample.

B. Microplate Procedure

1. Mix BCA Reagent A and BCA Reagent B in the ratio of 50:1. i.e., mix 50 ml of BCA Reagent A with 1 ml BCA Reagent B.

2. Follow Table 2 to prepare a fresh set of standards. (Dilute Albumin (BSA) Standards with 0.9% NaCl or PBS)

Table 2: Preparation of Albumin (BSA) Standards

Tube Number	Volume of Diluent (μl)	Volume of BSA (μl)	Final BSA Concentration (μg/ml)
A	0 μl	200 μl of 2 mg/ml Stock	2000 μg/ml
B	30 μl	90 μl of tube A	1500 μg/ml
C	60 μl	60 μl of tube A	1000 μg/ml
D	60 μl	60 μl of tube B	750 μg/ml
E	60 μl	60 μl of tube C	500 μg/ml
F	60 μl	60 μl of tube E	250 μg/ml
G	60 μl	60 μl of tube F	125 μg/ml
H	100 μl	25 μl of tube G	25 μg/ml
I	60 μl	0 0 (blank)	

3. Add 25 μl of each standard and protein samples into separate microplate wells.

4. Add 200 μl of BCA working reagent to each well and mix well.

5. Seal plates and incubate at 37°C for 30 minutes.

6. Cool plate to room temperature (RT).
7. Measure the absorbance at 562 nm on a plate reader within 10 minutes.
8. Subtract OD562 of Blank from all readings.
9. Plot the BSA standard curve: OD562 (on Y axis) vs BSA Standard concentration (on X axis). Use the standard curve to determine the protein concentration of each unknown sample.

Sensitivity	20 - 2000 µg/ml
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